

Farmer Name<sup>1</sup>

:Vet Name

Address:

Address:

Herd No:

Phone No.

Email:

Email:

**ELISA Test (antibody detection)<sup>2</sup>**

|  |  |                                     |   |                                   |                                     |
|--|--|-------------------------------------|---|-----------------------------------|-------------------------------------|
| <input type="checkbox"/> BVD p80                     | <input type="checkbox"/> Leptospira    | <input type="checkbox"/> Neospora   | <input type="checkbox"/> Chlamydia        | <input type="checkbox"/> Q-Fever  | <input type="checkbox"/> IBR gB     |
| <input type="checkbox"/> IBR gE                      | <input type="checkbox"/> Toxoplasma    | <input type="checkbox"/> Salmonella | <input type="checkbox"/> MAP              | <input type="checkbox"/> Fasciola | <input type="checkbox"/> Ostertagia |
| <input type="checkbox"/> BRSV                        | <input type="checkbox"/> M.haemolytica | <input type="checkbox"/> PI3        | <input type="checkbox"/> Mycoplasma bovis |                                   | <input type="checkbox"/> Pregnancy  |
| <input type="checkbox"/> IBR Tank Milk <sup>21</sup> |  |                                     |   |                                   |                                     |

 **ELISA Abortion package:** Neospora; Leptospira; Salmonella.

**PCR Test/Antigen ELISA Testing<sup>3</sup>**

|  |   |  |  |
|--|---|--|--|
| <input type="checkbox"/> BVD Virus PCR <sup>4</sup>  | <input type="checkbox"/> MAP PCR <sup>5</sup> | <input type="checkbox"/> Tick Borne Fever PCR <sup>6</sup><br><small>(EDTA blood samples only)</small> | <input type="checkbox"/> Mannheimia <sup>8</sup> |
| <input type="checkbox"/> Mycoplasma PCR <sup>9</sup> | <input type="checkbox"/> BVD Antigen ELISA    | <input type="checkbox"/> Histophilus Somni <sup>7</sup>  |  |

 Milk PCR<sup>10</sup>: *Staph. aureus; Strep. uberis, agalactiae, dysgalactiae; Mycoplasma bovis*
 Bovine abortion PCR<sup>11,12</sup>: Lepto, Chlamydia, Neospora

 Sheep abortion PCR<sup>11,12</sup>: Lepto, Chlamydia, Toxoplasma

 Respiratory virus PCR<sup>13</sup>: IBR, RSV, Coronavirus, PI3

**Parasitology<sup>14</sup>**

|  |  |                                   |                                 |
|--|--|-----------------------------------|---------------------------------|
| <input type="checkbox"/> Worm & Coccidia | <input type="checkbox"/> Liver fluke & | <input type="checkbox"/> Lungworm | <input type="checkbox"/> Faecal |
|--|--|-----------------------------------|---------------------------------|

**Milk Bacteriology:** Milk Culture<sup>15</sup>  Sensitivity testing<sup>16</sup>  Somatic Cell Count 
**Salmonella Culture**

**Calf Scour Package (E.coli,rota,corona,crypto)**  **Giardia** 
**Other Test:**<sup>17</sup>
**Sample Details**<sup>18</sup>

|   |                         |                        |                              |
|---|-------------------------|------------------------|------------------------------|
| Species:  | Sample Type:            | ICBF Y/N <sup>20</sup> | VetImpress Y/N <sup>20</sup> |
| Date Taken:   | History/clinical signs: |                        |                              |
| How do you wish samples to be tested? Individual/Pooled (Please delete one) |                         |                        |                              |
| <b>Animal ID:</b>   | <b>Tube code:</b>       | <b>Animal ID:</b>      | <b>Tube code:</b>            |
|   |                         |                        |                              |
|   |                         |                        |                              |
|   |                         |                        |                              |
|   |                         |                        |                              |
|   |                         |                        |                              |
|   |                         |                        |                              |
| Please add additional samples on a separate form                            |                         |                        |                              |

Before submitting samples, clients should ensure that they have read and understood FarmLab Diagnostics Terms and Conditions, and Sample packaging instructions available at [www.farmlab.ie](http://www.farmlab.ie).<sup>19</sup> **Explanatory Notes**

1. Animal ID and farmer herd number must be filled in for BVD and Johnes (MAP) testing. All positive results will be notified to the Department of Agriculture Food and the Marine in accordance with our terms and conditions.
2. Elisa antibody tests detect the presence of antibodies after an animal has been exposed to challenge either by natural infection, or in some cases, vaccination. The sensitivity and specificity of each test may vary. For more information please contact FarmLab Diagnostics
3. PCR tests detect the presence of DNA or RNA from the organism being tested for in the samples submitted. Positive results indicate that the organism is present in the sample. PCR tests will detect the presence of dead as well as living organisms. BVD antigen Elisa tests detect the presence of BVD virus in blood. This test is not suitable for use in calves less than 3 months old
4. BVD virus PCR tests detect the presence of BVDV RNA. The test may be carried out on blood, milk or ear notches. Samples may be pooled, however calves less than 60days of age must be tested individually.
5. MAP PCR detects the presence of Mycobacterium avium paratuberculosis DNA in faeces (Johnes Disease) . A positive results indicates that there is a high probability that the faeces contains MAP organisms. A negative result does not mean that the animal is free of Johnes Disease infection. This test is usually used as an ancillary test for animals which are suspected as being infected with MAP either on clinical signs, or as a result of a positive MAP elisa blood test.
6. Tick borne fever PCR detects the presence of *Anaplasma marginale* and *Anaplasma phagocytophilum*. Please submit individual blood samples in EDTA sampling tubes. Samples from up to 6 animals may be pooled in the laboratory to reduce the cost of testing
7. *Histophilus somni* PCR detects the presence of H.Somni DNA. It may be used on nasal swabs or lung tissue. H Somni is often a commensal in the upper respiratory tract
8. *Mannheimia haemolytica* PCR . Detects the presence of M.haemolytica serotypes most commonly associated with outbreaks of bovine respiratory disease.
9. *Mycoplasma* PCR detects the presence of *Mycoplasma bovis*. Suitable samples include milk, nasal swabs, joint fluid.
10. Milk PCR detects the presence of the organisms most commonly associated with infectious mastitis in cows. This test is suitable for use on individual and bulk tank samples. Positive results may be obtained even due to the presence of killed organisms.

11. Abortion PCR. This test detects the presence of organisms associated with abortion in cattle or sheep. The significance of positive chlamydia results in cattle is not fully understood. Suitable samples include placenta and foetal tissues (Kidney and brain) . A sample of blood should be submitted separately for brucellosis testing to the DAFM blood testing laboratory by the submitting veterinary surgeon.
12. **Abortion PCR tests should only be submitted on special FLOQswabs available on request from Farmlab**
13. Respiratory virus PCR. This test detects the presence of viruses associated with respiratory outbreaks in cattle. Submitting vets may also wish to consider including tests listed at 7,8 and 9 above. Plain cotton swabs which have been pre-moistened with saline should be used. Please ensure samples reach the laboratory as soon as possible. Where possible samples should be refrigerated before posting.
14. For further information on test interpretation please refer to our "Guide to the interpretation of parasitology results which accompanies all parasitology results.
15. Samples for milk culture should be taken aseptically from individual cows with clinical or subclinical mastitis. Udder quarters which are subclinically infected may be identified by using the somatic cell count test (17) or the California mastitis test. Standard milk culture is unsuitable for Bulk tank milk samples. For more information on BTM testing please contact the laboratory.
16. Sensitivity testing is carried out using antibiotic sensitivity discs on the predominant bacteria sp. which is identified on culture.
17. Other testing requests should be discussed with the laboratory before submission to clarify your testing requirements, and test availability. Requests for testing large numbers of samples (e.g greater than 500) should be notified to the laboratory in advance
18. Please fill in all elements of the "sample details" . **A valid tag number is required for all animals which are tested for BVDV and MAP. Tag numbers may be submitted separately on a printed list or excel file where it is not feasible to write all numbers on the submission form**
19. By submitting this form we assume that you have read our terms and conditions available on our website at . Samples must be packaged in accordance with our sample packaging instructions available on our website .
20. Please indicate by circling "Y" if you wish sample results to be transferred to either the ICBF or VetImpress databases
21. **We recommend using the whole IBR Tank Milk test for bulk tank milk samples due to higher sensitivity**