

Rev No: 18

| Vet Practice Na   | me                         |   | Client name     |                                       |                  |  |  |
|---|----------------------------|---|-----------------|---------------------------------------|------------------|--|--|
| Submitting vet  | name                       |   | Client herd no. |                                       |                  |  |  |
| Address   |                            |   | Address         | Address                               |                  |  |  |
|   |                            |   |                 |                                       |                  |  |  |
|   |                            |   |                 |                                       |                  |  |  |
| Practice email address  |                            |   |                 |                                       |                  |  |  |
| Practice phone number   |                            |   |                 |                                       |                  |  |  |
| Submitting vet mobile phone number  |                            |   |                 |                                       |                  |  |  |
| Sample Details <sup>18</sup>  |                            |   |                 |                                       |                  |  |  |
| CSV filename (for animal IDs supplied on a separate file)                                       |                            |   |                 |                                       |                  |  |  |
| TAL ACTH Voucher code <sup>(20,21)</sup>  |                            |   |                 |                                       |                  |  |  |
| Type of animal/ Brief History   |                            |   |                 |                                       |                  |  |  |
| Date of Sampling  |                            |   |                 |                                       |                  |  |  |
| Insert sample type: Blood (B) Individual Milk (IM) Bulk milk (BM) Individual faeces (IF) Pooled |                            |   |                 |                                       |                  |  |  |
| faeces (PF) Swal  | b (S) Tissue (1            | F). If other please specified in the specified of the second s | ecify.          |                                       |                  |  |  |
| the "comments"  | o de tested a:<br>7 column | s a pool, please indic  | ate which sampl | es are to be poo                      | bled together in |  |  |
| Animal ID/<br>Name  | Tube<br>Code               | Test Description  | Test<br>code    | Sample type:<br>B/IM/BM/IF/PF/<br>S/T | Comments         |  |  |
|   |                            |   |                 |                                       | _                |  |  |
|   |                            |   |                 |                                       | -                |  |  |
|   |                            |   |                 |                                       | -                |  |  |
|   |                            |   |                 |                                       | -                |  |  |
|   |                            |   |                 |                                       |                  |  |  |
| Table continued overleaf for multiple samples   |                            |   |                 |                                       |                  |  |  |
| ICBF (For AHI programmes)   |                            |   |                 |                                       |                  |  |  |
|   |                            |   |                 |                                       |                  |  |  |
| Submitted by: (Print Name) (Signature) (Date)   |                            |   |                 |                                       |                  |  |  |
|   |                            |   |                 |                                       |                  |  |  |

FarmLab Diagnostics have policies in place to ensure that data is maintained confidentially and in accordance with General Data Protection regulation requirements. Before submitting samples, clients should ensure that they have read and understood the following documents available at www.farmlab.ie/downloads

- FarmLab Diagnostics Terms and Conditions
- Policy on data protection
- Sample packaging instructions



## Multiple Samples

Multiple samples should be recorded in the table below. If numbers are supplied separately via csv file, the csv file name should be inserted in the Animal ID section above.

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| Tube | Animal ID | Test<br>Codo | Test Description | Sample type | Comments |
|------|-----------|--------------|------------------|-------------|----------|
| coue |           | Code         |                  |             |          |
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## F#514-1 FarmLab Diagnostics Sample Submission Form

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## **Explanatory Notes**

- 1. Animal ID and farmer herd number must be filled in for BVD and Johnes (MAP) testing. All positive results will be notified to the Department of Agriculture Food and the Marine in accordance with our terms and conditions.
- Elisa antibody tests detect the presence of antibodies after an animal has been exposed to challenge either by natural infection, or in some cases, vaccination. The sensitivity and specificity of each test may vary. For more information please contact FarmLab Diagnostics
- 3. PCR tests detect the presence of DNA or RNA from the organism being tested for in the samples submitted. Positive results indicate that the organism is present in the sample. PCR tests will detect the presence of dead as well as living organisms. BVD antigen Elisa tests detect the presence of BVD virus in blood. This test is not suitable for use in calves less than 3 months old
- 4. BVD virus PCR tests detect the presence of BVDV RNA. The test may be carried out on blood, milk or ear notches. Samples may be pooled, however calves less than 60days of age must be tested individually.
- 5. MAP PCR detects the presence of Mycobacterium avium paratuberculosis DNA in faeces (Johnes Disease). A positive results indicates that there is a high probability that the faeces contains MAP organisms. A negative result does not mean that the animal is free of Johnes Disease infection. This test is usually used as an ancillary test for animals which are suspected as being infected with MAP either on clinical signs, or as a result of a positive MAP elisa blood test.
- 6. Tick borne fever PCR detects the presence of Anaplasma marginale and Anaplasma phagocytophilum. Please submit individual blood samples in EDTA sampling tubes. Samples from up to 6 animals may be pooled in the laboratory to reduce the cost of testing
- 7. Histophilus somni PCR detects the presence of H.Somni DNA. It may be used on nasal swabs or lung tissue. H Somni is often a commensal in the upper respiratory tract
- 8. Mannheimia haemolytica PCR . Detects the presence of M.haemolytica serotypes most commonly associated with outbreaks of bovine respiratory disease.
- 9. Mycoplasma PCR detects the presence of Mycoplasma bovis. Suitable samples include milk, nasal swabs, joint fluid.
- 10. Milk PCR tests detect the presence of the organisms most commonly associated with infectious mastitis in cows. This test is suitable for use on individual or bulk tank samples. Positive results may be obtained even due to the presence of killed organisms.
- 11. Abortion PCR. This test detects the presence of organisms associated with abortion in cattle or sheep. The significance of positive chlamydia results in cattle is not fully understood. Suitable samples include placenta and foetal tissues(Kidney and brain). A sample of blood should be submitted separately for brucellosis testing to the DAFM blood testing laboratory by the submitting veterinary surgeon. Abortion PCR tests should only be submitted on special FLOQ swabs available on request from Farmlab
- 12. Respiratory virus PCR. This test detects the presence of viruses associated with respiratory outbreaks in cattle. Submitting vets may also wish to consider including tests listed at 7,8 and 9 above. Plain cotton swabs or swabs with viral transport medium should be used. Please ensure samples reach the laboratory as soon as possible. Where possible samples should be frozen before posting.
- 13. Parasitology: For further information on interpretation please refer to our "Guide to the interpretation of parasitology results which is available on our website.
- 14. Samples for milk culture should be taken aseptically from individual cows with clinical or subclinical mastitis. Udder quarters which are subclinically infected may be identified by using the somatic cell count test (17) or the California mastitis test. Standard milk culture is unsuitable for Bulk tank milk samples. For more information on BTM testing please contact the laboratory. Sensitivity testing is carried out using antibiotic sensitivity discs on the predominant bacteria sp. which is identified on culture.
- 15. We recommend using the whole IBR Tank Milk test for bulk tank milk samples due to higher sensitivity. This test is suitable for use on pools of up to 128 animals. It is recommended to use the test as a monitoring tool and repeat 3-4 times yearly. A negative result does not indicate freedom from disease, herds with a low, less than 10%, within herd prevalence may yield a negative result. Where bulk tank milk samples are submitted for IBRgB testing, we will also test using the IBR Bulk Tank Milk elisa unless requested not to do so. Samples submitted from vaccinated herds being submitted for IBRgE testing will just be tested using the IBRgE milk elisa. Clients should note that the sensitivity of this test on pooled milk samples is low, requiring a minimum of 20% of the herd to be positive before the test will yield a positive result.
- 16. Other testing requests should be discussed with the laboratory before submission to clarify your testing requirements, and test availability. Requests for testing large numbers of samples (e.g greater than 500) should be notified to the laboratory in advance
- 17. Please fill in all elements of the "sample details". A valid tag number is required for all animals which are tested for BVDV and MAP. Tag numbers may be submitted separately on a printed list or excel file where it is not feasible to write all numbers on the submission form
- 18. By submitting this form we assume that you have read our terms and conditions available on our website at. Samples must be packaged in accordance with our sample packaging instructions available on our website .
- 19. CEM swabs must be put on test within 48hrs of sampling. Where samples do not arrive in time to ensure this takes place a resample will be requested. Samples yielding a heavily contaminated culture growth may also have to be re-submitted. Complete Identification details should be submitted for all CEM samples. Details of CEM sample results will be shared with the Department of Agriculture
- 20. Samples for ACTH analysis should be collected in EDTA tubes and should reach the laboratory within 24hrs of the samples being taken. Ideally samples should be centrifuged and the plasma separated. Samples should be kept cool during shipping by the use of cool packs.
- 21. ACTH samples submitted under the Talk about Laminitis (TAL) scheme provided by Boehringer Ingelheim Animal Health UK Ltd should be submitted with a TAL voucher code generated from www.talkaboutlaminitis.ie. Voucher codes cannot be backdated. For more information visit www.talkaboutlaminitis.ie or contact your Boehringer Ingelheim Territory Manager.

|                            | F#514-1 FarmLab Diagnostics Sample Submission Form |
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| <b>FarmLab DIAGNOSTICS</b> | Rev No: 18   |